

Specification

General purpose solid medium containing animal and plant peptone, and neutralisers, according to Pharmacopoeial Harmonised Method and ISO Standards.

Presentation

	Packaging Details	Shelf Life	Storage
30 Contact Plates/lrd Contact Plates - Triple Wrapping with: 15 ± 2 ml	1 box with 3 x 10 plates BOPP plastic bags (triple wrapping) with stacks of 5 plates inside. Every pack exhibits a irradiation indicator stacked on the side of the bag (8-14 KGy). LATERAL LABELLING LOCKABLE PETRI LID	9 months	15-25 °C

Composition

Composition (g/l):	
Peptone from casein	15.0
Soya peptone.....	5.00
Sodium chloride.....	5.00
Histidin.....	1.00
Lecithine.....	0.70
Polysorbate 80.....	5.00
Sodium Thiosulfate.....	0.50
Agar.....	15.0

Description /Technique

Description

TSA is a widely used medium containing two peptones which support the growth of a wide variety of organisms, even that of very fastidious ones such as Neisseria, Listeria, Brucella, etc. It is frequently used for routine diagnostic purposes due to its reliability and its easily reproducible results.

The addition of the neutralizing agents TLHTh (Tween 80 - Lecithin - Histidine - Sodium Thiosulphate) may inactivate a variety of disinfectants.

- * The combination of lecithin, polysorbate 80 and histidine neutralizes aldehydes and phenolic compounds.
- * The combination of lecithin and polysorbate 80 neutralizes the quaternary ammonium compounds.
- * The polysorbate 80 neutralizes hexachlorophene and mercurial derivatives.
- * Sodium thiosulphate neutralizes halogen compounds.
- * Lecithin neutralizes chlorhexidine.
- * Histidine neutralizes formaldehyde.

Technique

Collect, dilute and prepare samples and volumes as required according to specifications, directives, official standard regulations and/or expected results.

This medium is also well suited for air environmental sampling (total compatibility with most commercially available air samplers) or for other types of environmental sampling (fingers or gloves of operators, swab streaking,...).

Spread the plates by streaking methodology or by spiral method.

The inoculated plates are incubated at 30-35 ° C for 24-72 h (bacteria) and 3-5 days for fungi (yeast & molds). Examined daily.

(Incubation times greater than those mentioned above or different incubation temperatures may be required depending on the sample, on the specifications,... This medium can be inoculated directly or after enrichment broth).

After incubation, enumerate all the colonies that have appeared onto the surface of the agar.

Each laboratory must evaluate the results according to their specifications.

Calculate total microbial count per ml of sample by multiplying the average number of colonies per plate by the inverse dilution factor if streaked a diluted sample. Report results as Colony Forming Unit (CFU's) per ml or g along with incubation time and temperature.

The lid can be used locking the plate in two positions after taking the sample:

- **AIR:** lid closed, but leaving certain movement, for AEROBIC and ANAEROBIC incubations.
- **CLOSE:** lid completely closed. Better for transport, avoiding risk of contamination due to its possible opening during the transport.

Attention: Plates are used for monitoring the microbiological contamination of surface and air inside cleanrooms, isolators, RABS, food industries and hospitals. The double/triple irradiated wrapping ensures that the package itself doesn't contaminate the environment as the first wrapper is removed just before entering the clean area.

Wrapping resistant to hydrogen peroxide vapors penetration.

Quality control
Physical/Chemical control

Color : Straw-coloured yellow pH: 7.3 ± 0.2 at 25°C

Microbiological control

Growth Promotion Test 50-100 CFU according to harmonized Pharmacopoeia monographs (EP) and test methods & ISO 11133:2014/A1:2018
Analytical methodology according to ISO 11133:2014/A1:2018; A2:2020.

Aerobiosis. Incubation at 30-35-37 °C. Read after 18-24 h to 72 h for bacteria and 3-5 days for fungi.

Microorganism
Growth

<i>Escherichia coli</i> ATCC® 8739, WDCM 00012	Good (≥70%)
<i>Staphylococcus aureus</i> ATCC® 6538, WDCM 00032	Good (≥70%)
<i>Bacillus subtilis</i> ATCC® 6633, WDCM 00003	Good (≥70%)
<i>Candida albicans</i> ATCC® 10231, WDCM 00054	Good (≥70%)
<i>Ps. aeruginosa</i> ATCC® 9027, WDCM 00026	Good (≥70%)
<i>Salmonella typhimurium</i> ATCC® 14028, WDCM 00031	Good (≥70%)
<i>Aspergillus brasiliensis</i> ATCC® 16404, WDCM 00053	Good (≥70%)
<i>L. monocytogenes</i> ATCC® 13932, WDCM 00021	Good (≥70%)
<i>Bacillus cereus</i> ATCC® 11778, WDCM 00001	Good (≥70%)
<i>Enterococcus faecalis</i> ATCC® 29212, WDCM 00087	Good (≥70%)
<i>Clostridium perfringens</i> ATCC® 13124, WDCM 00007	Good (≥70%)
<i>Clostridium sporogenes</i> ATCC® 19404, WDCM 00008	Good (≥70%)
<i>Stph. aureus</i> ATCC® 25923, WDCM 00034	Good (≥70%)
<i>Escherichia coli</i> ATCC® 11775, WDCM 00090	Good (≥70%)

Sterility control

Incubation 48 h at 30-35 °C and 48 h at 20-25 °C: NO GROWTH.
Check at 7 days after incubation in same conditions.

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