

Specification

Solid and selective medium for the isolation of yeasts and moulds from the environment and food products.

Presentation

	Packaging Details	Shelf Life	Storage
30 Contact Plates/lrd. Contact Plates - Double Wrapping with: 15 ± 2 ml	1 box with 5 blisters (PET laminated and PPBO bag) with 6 contact plates/blister. Every pack exhibits an irradiation indicator (8-14kGy).	5 months	2-25 °C

Composition

Composition (g/l):	
Mycological peptone.....	5.000
Dextrose.....	10.000
Potassium phosphate.....	1.000
Magnesium sulfate.....	0.500
Rose bengal.....	0.050
Chloramphenicol.....	0.100
Agar.....	15.000

Description /Technique

Description:

Rose Bengal Agar is a selective medium used to detect and enumerate moulds and yeasts in food samples. In addition the nutritional requirements for moulds and yeasts, this medium also contains Rose Bengal, which apart from turning the yeast a pink colour, facilitates counting, by reducing the luxuriant growth of moulds such as Rhizopus and Neurospora. This makes it is easier to detect other slower growing moulds.

The chloramphenicol included in the formulation inhibits bacterial growth, but does not interfere with the growth of fungi.

Technique

Contact plates are used in the microbiological control of disinfection and cleaning of surfaces. It acts simultaneously as a sampler and incubation culture medium without the need for any other intermediate steps.

The plates come in a form appropriate for this function and can be used with different culture media depending on the type of microbe that needs to be controlled. On average the plates provide a contact surface of approximately 25 cm².

To use, remove the cover and gently press the culture medium on the surface to be controlled, ensuring contact between the two surfaces. The Contact plate is removed and covered with the lid to prevent air contamination. It is advisable that the lid is secured with adhesive tape and the bottom labelled with the sampling data (place, date and time). The inoculated plates are incubated at 25 ±1 ° C for 5 days and examined daily.

If the sample surfaces are rough, the Contact plates will not make good contact, even when the pressure is increased. In these cases it is advisable to delineate an sample surface area of 25 cm squared and rub this area vigorously with a wet sterile swab and then rub the swab over the Contact plate.

If verifying the effectiveness of a cleaning or disinfection process, Contact plates should be used within two hours after the end of the process, ensuring that the sample surface is dry. It is advisable to always include positive controls, sampling the area before disinfection or dirty areas beside the disinfected area.

The technician will determine the frequency of sampling and disinfection according to performance criteria.

Apply the agar directly onto surface to be monitored ensuring that the pressure is distributed over the whole plate for 10 seconds.

Note: Contact plates are used for monitoring the microbiological contamination of surface and air inside cleanrooms, isolators, RABS, food industries and hospitals. The double/triple irradiated wrapping ensures that the package itself doesn't contaminate the environment as the first wrapper is removed just before entering the clean area.

The plates must be kept in their original packaging (blisters) to guarantee their stability at the end of their expiration date.

Quality control**Physical/Chemical control**

Color : Strongly pink

pH: 7.2 ± 0.2 at 25°C

Microbiological controlInoculate: Practical range 100 ± 20 CFU. min. 50 CFU (productivity)/ 10⁴-10⁶ (selectivity).

Microbiological control according to ISO 11133:2014/A1:2018.

Analytical methodology according to ISO 11133:2014/A1:2018; A2:2020.

Aerobiosis. Incubation at 25 °C ±1, reading at 72 h to 5 days.

Microorganism*Candida albicans* ATCC® 10231, WDCM 00054*Aspergillus brasiliensis* ATCC® 16404, WDCM 00053*Escherichia coli* ATCC® 8739, WDCM 00012*Bacillus subtilis* ATCC® 6633, WDCM 00003**Growth**

Good (≥50 %)

Good (≥50 %)

Inhibited

Inhibited

Sterility control

Incubation 48 h at 30-35 °C and 48 h at 20-25 °C: NO GROWTH.

Check at 7 days after incubation in same conditions.

Bibliography

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