

Specification

Sterile selective supplement for the isolation of *Legionella* spp. from environmental water samples.

Presentation

	Packaging Details	Shelf Life	Storage
10 Freeze dried vials			
Vial	22±0.25 x 55±0.5 mm glass vials, tag labelled, White	49 months	2-25 °C
with: 3 ± 0.1 g	plastic cap - 10 vials per box.		

Composition

Composition (vial)

Polymyxin B sulfate.....	40000,00 IU
Sodium cefazolin.....	4,50 mg
Pimaricin (syn Natamycin).....	35,00 mg

Reconstitute the original freeze-dried vial

by adding :

Sterile Distilled Water..... 9 ml

Description /Technique

Description:

The discovery of the causative organism of Legionnaires' disease has permitted big progress in the studies around it. New media for the culture and the enumeration *Legionella* spp have been developed in the last years.

Legionella BCYE + AB selective supplement, when added to the agar Base, gives the antibiotic support in order to obtain a selective final medium.

The selectivity is raised by the addition on sodium cefazolin that acts against Gram positive bacteria, polymyxin B that inhibits Gram negative bacteria and natamycin that are antifungal agents and inhibits the yeast growth.

Technique:

Collect, dilute and prepare samples and volumes as required according to specifications, directives, official standard regulations and/or expected results.

Reconstitute the 1 vial of Selective Supplement of *Legionella* BCYE + AB with 9 ml to the steril distilled water, in aseptic conditions, and add it to 500 ml of melted Legionella BCYE Agar Base cooled to 47- 50°C supplemented before with Legionella BCYE growth Supplement. Do not overheat once supplemented.

Pour the complete medium into Petri dishes and, once solidified on a flat surface. Spread the plates by streaking methodology or by MF method.

Allow the inoculated plates to stand until the inocula has been absorbed. Invert the plates and incubate at 36 ± 2°C for up to 2, 3, 5 -10 days. To ensure the atmosphere in the incubator is humid, place a tray of water in the bottom of the incubator. Top up this tray with fresh water (if necessary) each time the plates are examined. Incubation in an atmosphere of air with 2,5% (volume fraction) CO₂ may be beneficial for the growth of some *Legionella*, but it is not essential.

Examine the plates with a plate microscope on at least three occasions at intervals of 2,3 to 5 days during the 10-day incubation period, as *Legionella* grow slowly and can be masked by the growth of other organisms. Record the number of each type of colony present. Colonies of *Legionella* are often white-grey-blue-purple in colour, but may be brown, pink, lime-green or deep-red. They are smooth with a smooth edges and exhibit a characteristic ground-glass appearance. Under ultraviolet light colonies of several species autofluoresce brilliant white, but others are red and *L. pneumophila* appear dull green often tinged with yellow. All presumptive colonies must be confirmed by cultural, biochemical, serological or genetic methods.

Quality control

Physical/Chemical control

Color : White

Microbiological control

Reconstitute 1 vial as indicated in COMPOSITION; shake and dissolve completely

Add 1 vial to 500 ml of medium base. DO NOT HEAT once supplemented.

Aerobiosis. Incubation at 36 ± 2 °C. Reading 3 - 5 days, up to 10 days.

Microorganism

Escherichia coli ATCC® 8739, WDCM 00012

Enterococcus faecalis ATCC® 19433, WDCM 00009

L. anisa ATCC® 35292, WDCM 00106 (by MF)

L. pneumophila ATCC® 33152, WDCM 00107 (by MF)

the reference medium is GVPC validated.

Growth

Inhibited

Inhibited

Good ($\geq 50\%$) grey-blue colonies

Good ($\geq 50\%$) grey-blue colonies

Sterility control

Add 5 ml of the sample to:

100 ml TSB and 100 ml Thioglycollate.

Incubation 48 hours at 30-35 °C and 48 hours at 20-25 °C: NO GROWTH.

Bibliography

- ATLAS, R.M. & L.C. PARKS (1993) Handbook of Microbiological Media. CRC Press. BocaRaton. Fla. USA.
- CLESCERI, L.S., A.E. GREENBERG & A.D. EATON (1998) Standard methods for the examination of water and wastewater. 9-106. 20th edition. APHA-AWWA-WEF. Washington DF, USA.
- EDELSTEIN, P.H., (1981) Improved semiselective medium for the isolation of *Legionella pneumoniae* from contaminated clinical and environmental specimens. J. Clin Microbiol. 14(3):298.
- FEELEY, J.C., R.J. GIBSON, G.W. GORMAN, N.C. LANGFORD, J.K. RASHEED, C.D. MACKEL, & W.B. BAINE (1979) Charcoal-Yeast Extract Agar: Primary isolation medium for *Legionella pneumophila*. J. Clin. Microbiol. 10(4) 437.
- ISO 11731 Standard (2017) Water Quality - Enumeration of *Legionella*.
- ISO 11133:2014/ Adm 1:2018/ Adm1 :2018. Microbiology of food, animal feed and water. Preparation, production, storage and performance testing of culture media.
- MacFADDIN, J.F. (1985) Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria.
- PASCULLE, A.W., J.C. FEELEY, R.J. GIBSON, L.G. CORDES, R.L. MYEROWITZ, C.M. PATTON, G.W. GORMAN, C.L. CARMACK, J.W. EZZELL & J.N. DOWLING (1980) Pittsburgh pneumonia agent: Direct isolation from human lung tissue. J. Infect. Dis., 141:727.
- UNE-EN ISO 11133 (2014). Microbiología de los alimentos para consumo humano, alimentación animal y agua.-Preparación, producción, conservación y ensayos de rendimiento de los medios de cultivo.
- WARD, K.W. (1995) Processing and interpretation of specimens for *Legionella spp*. In "Clinical Microbiology Procedures Handbook" Chap. 12.1 edited b H.D. Isenberg. ASM Press. Washington DC, USA.