

**Product :**
**CHARCOAL CEFOPERAZONE  
DEOXYCHOLATE MODIFIED AGAR BASE (M-  
CCDA)**
**Also known as**

mCCDA

**Specification**

Selective plating medium used for the detection and enumeration of *Campylobacter spp* according to the ISO 10272 standard.

**Formula \* in g/L**

Meat extract.....	10,00
Peptone.....	10,00
Sodium chloride.....	5,00
Bacteriological charcoal.....	4,00
Casein hidrolysate.....	3,00
Sodium deoxycholate.....	1,00
Iron (II) sulfate.....	0,25
Sodium pyruvate.....	0,25
Agar.....	15,00

Final pH 7,4 ± 0,2 at 25 °C

\* Adjusted and /or supplemented as required to meet performance criteria

**Directions**

Suspend 24.2 g of powder in 500 mL of distilled water and bring to the boil to dissolve. Sterilize in an autoclave at 121°C for 15 minutes. Cool to 47-50°C and aseptically add one vial of the *Campylobacter* CCDA Selective Supplement (Art. No. DSHB3110). Mix carefully and pour into sterile Petri dishes.

Note: If the plates are prepared in advance, they should be kept for not more than 4 hours at ambient temperature or for no more than 7 days in the dark at 3 ± 2°C.

**Description**

CCD Modified Agar is formulated according to the ISO Standard 10272-1:2006 and is intended to detect and enumerate *Campylobacter spp* from food and animal feeding stuffs.

After determining that *Campylobacter* species grow best on solidified Nutrient Broth No. 2 compared to other media workers (1983) carried out a systematic survey of alternatives to blood for neutralizing oxygen toxicity. A combination of 0,4% charcoal, 0,25% ferrous sulfate and 0,25% sodium pyruvate proved best.

A further study surveyed the suppressive effect of several inhibitors on the undesirable microbiota showing deoxycholate and cefazolin as the most effective inhibitory agents. Later, in 1984, Hutchinson and Bolton replaced cefazolin (10 mg/L) with cefoperazone (32 mg/L). This allowed fewer contaminants to grow, and permitted the modified medium (modified CCD Agar or mCCDA) to be used at 37°C. However amphotericin B was needed to prevent overgrowth by yeast able to grow at 37°C but not at 41,5 ± 1°C.

In 1993 Aspinall *et al.* developed a modification of mCCDA designed for use at 37°C to isolate *C. upsaliensis* as well as the other thermophilic *campylobacter* species. This medium contains 8 mg/L cefoperazone and 4 mg/L teicoplanin replacing 32 mg/L cefoperazone in mCCDA. Teicoplanin has an antimicrobial spectrum similar to that of vancomycin, active mainly against Gram positive bacteria. By comparison with mCCDA the final formulation of this medium, called CAT Agar, isolated the same numbers of *Campylobacter spp* other than *C. upsaliensis* from faeces and is superior to mCCDA for *C. upsaliensis* with slightly higher growth of competing microflora.

**Necessary supplements**

*Campylobacter* CCDA Selective Supplement (Art. No. DSHB3110)

Vial Contents:

Necessary amount for 500 mL of complete medium.

Amphotericin B..... 5,00 mg

Cefoperazone..... 16,00 mg

Distilled water (Solvent)

**Product :**
**CHARCOAL CEFOPERAZONE  
DEOXYCHOLATE MODIFIED AGAR BASE (M-  
CCDA)**
**Technique**

Immediately before use, carefully dry the agar plates, preferably with the lids off and the agar surface downwards, in a drying cabinet, until the agar surface is free of visible moisture (maximum 30 minutes).

Using the culture obtained from enrichment broth (Bolton Broth), inoculate the mCCDA with a sterile loop. Incubate plates at 41,5°C in a microaerobic atmosphere (approximately 5% O<sub>2</sub>, 10% CO<sub>2</sub> and 85% N<sub>2</sub> or H<sub>2</sub>), for 44 ± 4 hours.

- *Campylobacter jejuni* strains produce grey, moist flat and occasionally spreading growth which may be accompanied with a green hue and/or a metallic sheen.
- *Campylobacter coli* strains tend to be creamy-grey in colour, moist and often produce a more discrete type of colony.
- *Campylobacter lari* strains are more varied and produce both types of colonial morphology.
- Occasionally contaminating organisms may grow on this medium. These include cefoperazone-resistant *Pseudomonas* spp, Enterobacteriaceae, and some streptococci and yeasts.

**Quality control**

**Incubation temperature:** 41,5 ± 1°C

**Incubation time:** 44 ± 4h

**Inoculum:** Practical range 100 ± 20 CFU. Min. 50 CFU (productivity) / 10<sup>4</sup>-10<sup>6</sup> CFU (selectivity) according to ISO 11133:2014/Amd 1:2018 .

**Microorganism**
**Growth**
**Remarks**

*Campylobacter jejuni* ATCC® 29428

Productivity > 0.50

Under microaerophilic atmosphere

*Campylobacter coli* ATCC® 43478

Productivity > 0.50

Under microaerophilic atmosphere

*Escherichia coli* ATCC® 8739

Partial inhibition

-

*Staphylococcus aureus* ATCC® 25923

Total inhibition

-

**References**

- ASPINALL, S.T., D.R.A. WAREING, P.G. HAYWARD & D.N. HUTCHINSON (1993) Selective medium for thermophilic campylobacters including *Campylobacter upsaliensis*. J. Clin. Pathol. 46:829-831.
- BAYLIS, C.L., (editor) (2007) Manual of Microbiological Methods for the Food and Drinks Industry. 5th Edition Method 3.3.1:2007. CCFRA .Chipping Campden. U.K.
- BOLTON, F.J. (2000) Methods for isolation of campylobacters from humans, animals, food and water. In "The increasing incidence of human campylobacteriosis" Report and Proceedings of a WHO Consultation of Experts. Copenhagen Denmark 21-25 November 2000, WHO/CDS/ CSRAPH 2001. p. 87-93.
- BOLTON, F.J., D. COATES, (1983) Development of a blood-free campylobacter medium: screening tests on basal media and supplements, and the ability of selected supplements to facilitate aerotolerance. J. Appl. Bacteriol. 54:115-125.
- BOLTON, F.J., D. COATES & D.N. HUTCHINSON (1984) The ability of *Campylobacter* media supplements to neutralize photochemically induced toxicity and hydrogen peroxide. J. Appl. Bacteriol. 56:151-157.
- CORRY, J.E.L., H. IBRAHIM ATABAY, S.J. FORSYTHE & L.P. MANSFIELD (2003) Culture Media for the isolation of campylobacters, helicobacters and arcobacters. In Handbook of Culture Media for Food Microbiologists. J.E.L. Corry et al. (Eds.) Elsevier Science B.V. Amsterdam.
- CORRY, J.E.L., G.D.W. CURTIS & R.M. BAIRD (2003) Handbook of culture media for food Microbiology. Elsevier Sci. B. V. Amsterdam.
- FDA (Food and Drug Administrations) (1998) Bacteriological Analytical Manual. 8th Edition. Revision A. AOAC International. Gaithersburg, Maryland, USA.
- HUNT, J.M., C. ABEYTA & T. TRAN (1998) *Campylobacter*. In FDA BAM 8th Edition (revision A) 7.01-7.027 AOAC International. Gaithersburg, Md, USA.
- HUTCHINSON, D.N. & F.J. BOLTON (1984) Improved blood-free selective medium for the isolation of *Campylobacter jejuni* from faecal specimens. J. Clin Pathol. 37:956-957.
- ISO 10272-1 Standard (2017) Microbiology of the food chain - Horizontal Method for detection and enumeration of *Campylobacter* spp. - Part 1: Detection method.
- ISO 10272-2 Standard (2017) Microbiology of the food chain - Horizontal Method for detection and enumeration of *Campylobacter* spp. - Part 2: Colony count-technique.
- ISO 11133:2014/ Adm 1:2018. Microbiology of food, animal feed and water. Preparation, production, storage and performance testing of culture media.
- STERN, N.J., J.E. LINE & H.C. CHEN (2001) *Campylobacter* In "Compendium of methods for the Microbiological Examination of Foods" 4th Ed. F.P. Downes & K. Ito (Eds.) APHA, Washington DC. USA.

**Storage**

For laboratory use only. Keep tightly closed, away from bright light, in a cool dry place (+4 °C to 30 °C).