


**Formula \* in g/L**

Proteose peptone.....	10.000
Yeast extract.....	10.000
Sodium chloride.....	5.000
Sodium glycerophosphate.....	10.000
Maltose.....	20.000
Lactose.....	1.000
Sodium azide.....	0.400
Bromocresol purple.....	0.015
Agar.....	20.000

Final pH 7,2 ±0,2 at 25 °C

\* Adjusted and /or supplemented as required to meet performance criteria

**Directions**

Suspend 76.4 g of powder in 1 L of distilled water and bring to the boil stirring constantly. If it is to be used immediately, sterilisation is not necessary. If sterilization is necessary, sterilize in the autoclave in small volumes, at 121°C for 10 minutes maximum. Let agar cool to 50°C and add 10 mL/L of TTC Sterile Solution 1% (Art. No. DSHB3074). Homogenize well and distribute into Petri dishes.

Note: A non-homogeneous appearance is normal, and does not affect the medium's quality and efficacy.

**Description**

Kenner, Clark and Kabler (1960, 1961) discovered that KF medium was excellent for detecting enterococci in polluted water. Carbohydrates in this medium (lactose and maltose) are utilised by most of enterococci, producing a big amount of acid and causing the indicator to turn from violet to yellow. Streptococci that do not belong the D group may also grow in the medium, but they do not produce enough acid to change the colour of the indicator. Other microorganisms are strongly inhibited by sodium azide. Enterococci reduce TTC to formazan and so their colonies are red in colour.

**Technique**

If the sample is suspected of being highly contaminated, prepare ten-fold serial dilutions bank and inoculate the surface with 0,1 mL of sample using a Drigalsky loop (spread plate method) or, if desired, inoculate the medium with 1 mL of sample using the pour plate method. Incubation should be carried out at 37°C for a 48 hours period.

After incubation, readings are performed by observing whether the indicator has turned from violet to yellow, and whether colonies are pink or red in colour.

It is very important to maintain the pH of the medium over 7,0, or otherwise, false results may occur. Sterilization for longer than the specified period could result in darkening of the sugar and thereby resulting in a decrease in the pH.

**Quality control**

**Incubation temperature:** 37°C ± 1

**Incubation time:** 48 h ± 3

**Inoculum:** Practical range 100 ± 20 CFU. Min. 50 CFU (Productivity) /10<sup>4</sup>-10<sup>6</sup> CFU (Selectivity). Spiral Plate Method (or Membran Filter Method).

**Microorganism**
**Growth**
**Remarks**

*Pseudomonas aeruginosa* ATCC® 27853

Total inhibition

Selectivity

*Escherichia coli* ATCC® 25922

Total inhibition

Selectivity

*Enterococcus faecalis* ATCC® 29212

Productivity > 0.50

Dark red colonies.Yellowish medium

*Enterococcus faecalis* ATCC® 19433

Productivity > 0.50

Dark red colonies.Yellowish medium

**References**

- CLESCERI, L.S., A.E. GREENBERG & A.D. EATON (1998) Standard Methods for the Examination of Water and Wastewater 20th ed. APHA, Washington.
- DOWNES, F.P. & K. ITO (2001) Compendium of Methods for the Microbiological Examination of Foods. 4th ed. APHA. Washington.
- KENNER, B.A., CLARK, H.F. & KABLER, P.W. (1960) Fecal Streptococci I. Cultivation and Enumeration of Streptococci in Surface Waters. Appl. Microbiol. 9:15.
- KENNER, B.A., CLARK, H.F. & KABLER, P.W. (1961) Fecal Streptococci II. Quantification of Streptococci in faeces. Am. Inst. Publ. Health, 50:1553.

**Storage**

For laboratory use only. Keep tightly closed, away from bright light, in a cool dry place (+4 °C to 30 °C).